EXPECTED VALUES AND SENSITIVITY
Healthy individuals are expected to have B2MG serum or plasma values 0 - 2.0 µg/ml and urine values 0 – 0.3 µg/ml. The minimum detectable sensitivity is estimated to be 0.1 µg/ml.

REFERENCES

2008-01-15
(Mfg.: 040203)

Warning

All GenWay kits have not been tested for clinical use and are not approved in the United States by the FDA for diagnostic clinical use. They are components or reagents made solely for research use, further manufacturing and export use. It is the commitment of GenWay customers to receive its products solely for the purpose of exportation or research, and not for the purposes of clinical diagnostic use.

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INTENDED USE
The is an enzyme immunoassay for the quantitative determination of Beta-2 Microglobulin (B2MG) Concentration in Human Serum, Plasma and Urine. Its common name is 3-2 MG Enzyme Immunoassay. For research use only.

SUMMARY AND EXPLANATION
Human 3-2 Microglobulin (B2MG) is an 11.8 kD protein identical to the light chain of the HLA-A, -B, and -C antigen. B2MG is expressed on nucleated cells, and is found at low levels in the serum and urine of normal individuals. B2MG concentrations are increased in inflammatory diseases, some viral diseases, renal dysfunction, and autoimmune diseases. A number of publications are available which explain the interpretation of B2MG serum levels in assessing the status of individuals with various clinical conditions.

PRINCIPLE OF THE TEST
The B2MG ELISA test is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay system utilizes a unique monoclonal antibody directed against a distinct antigenic determinant on the intact 3-2 Microglobulin molecule. Mouse monoclonal anti-B2MG antibody is used for solid phase immobilization (on the microtiter wells). A sheep anti-B2MG antibody is in the antibody-enzyme (horseradish peroxidase) conjugate solution. The diluted test sample is allowed to react first with the immobilized antibody for 30 minutes at 37°C. The sheep anti-B2MG-HRP conjugate is then added and reacted with the immobilized antigen for 30 minutes at 37°C, resulting in the B2MG molecules being sandwiched between the solid phase and enzyme-linked antibodies. The wells are washed with water to remove unbound-labeled antibodies. A solution of TMB Reagent is added and incubated for 20 minutes at room temperature, resulting in the development of a blue color. The color development is stopped with the addition of Stop Solution, changing the color to yellow. The concentration of B2MG is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.

MATERIALS PROVIDED

<table>
<thead>
<tr>
<th>MATERIALS PROVIDED</th>
<th>96 TESTS</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Microwells coated Murine monoclonal anti-B2 MG antibody</td>
<td>12x8x1</td>
</tr>
<tr>
<td>2. B2MG Reference Standards: 0, 0.625, 1.25, 2.5, 5, and 10</td>
<td>0.625 µg/ml</td>
</tr>
<tr>
<td>3. Sample Diluent, 100 ml</td>
<td>100 ml</td>
</tr>
<tr>
<td>4. Enzyme Conjugate Reagent, 22 ml</td>
<td>22 ml</td>
</tr>
<tr>
<td>5. TMB Reagent (One-Step), 11 ml</td>
<td>11 ml</td>
</tr>
<tr>
<td>6. Stop Solution (1N HCl), 11 ml</td>
<td>11 ml</td>
</tr>
</tbody>
</table>

MATERIALS NOT PROVIDED

1. Distilled or deionized water
2. Precision pipettes
3. Disposable pipette tips
4. ELISA reader capable of reading absorbance at 450nm
5. Absorbance paper or paper towel
6. Graph paper

STORAGE AND STABILITY

1. Store the kit at 2-8°C.
2. Keep microwells sealed in a dry bag with desiccants.
3. The reagents are stable until expiration of the kit.
4. Do not expose test reagents to heat, sun or strong light.
WARRIORS AND PRECAUTIONS

1. Potential biohazardous materials:
   The calibrator and controls contain human source components which have been tested and found non-reactive for hepatitis B surface antigen as well as HIV antibody with FDA licensed reagents. However, there is no test method that can offer complete assurance that HIV, Hepatitis B virus or other infectious agents are absent. These reagents should be handled at the Biosafety Level 2, as recommended in the Centers for Disease Control/National Institutes of Health manual, "Biosafety in Microbiological and Biomedical Laboratories" 1984.

2. This kit is designed for Research Use only.

3. Optimal results will be obtained by strict adherence to the test protocol. Precise pipetting as well as following the exact time and temperature requirements is essential.

4. Do not pipette by mouth. Do not smoke, eat, or drink in the areas in which specimens or kit reagents are handled.

5. The components in this kit are intended for use as an integral unit. The components of different lots should not be mixed.

6. Control sera and sample diluent contain preserved with sodium azide. Sodium azide may react with lead and copper plumbing to form explosive metal azide. On disposal, flush with a large volume of water.

SPECIMEN COLLECTION AND HANDLING

1. Blood should be drawn using standard venipuncture techniques and the serum should be separated from the red blood cells as soon as practical. Avoid grossly hemolytic, lipemic or turbid samples.

2. Specimens should be capped and may be stored for up to 48 hour at 2-8°C prior to assaying. Specimens held for a longer time can be frozen at -20°C for up to 6 months prior to assay. Thawed specimens should be inverted several times to mix prior to testing.

3. Collect urine samples and store at 2-8°C for up to 5 days or at -20°C for longer periods. Urine samples are diluted 1:10 by adding 50 μl urine to 450μl sample diluent. Use same assay procedure as for serum test.

PREPARATION FOR ASSAY

1. All reagents should be brought to room temperature (18-26°C) before use. All reagents should be mixed by gently inverting or swirling prior to use. Do not induce foaming.

2. Reconstitute each lyophilized standard with 1.0 ml distilled water. Allow the reconstituted material to stand for at least 20 minutes and mix gently. Reconstituted standards will be stable for up to 30 days when stored sealed at 2-8°C.

ASSAY PROCEDURE FOR SERUM AND PLASMA

1. Samples of patient serum, plasma and control serum need to be diluted before use for best results. Prepare a series of small tubes (such as 1.5 ml microcentrifuge tubes) and mix 10 μl serum with 1.0 ml Sample Diluent (101 fold dilution). Do not dilute the standards, they have already been pre-diluted 101 fold.

2. Secure the desired number of coated wells in the holder.

3. Dispense 20 μl of standards, diluted specimens, and diluted controls into appropriate wells.

4. Dispense 200 μl of Sample Diluent into each well.

5. Thoroughly mix for 30 seconds. It is very important to mix them completely.

6. Incubate at 37°C for 30 minutes.

7. Remove the incubation mixture by flicking plate contents into a waste container.

8. Rinse and flick the microtiter wells 5 times with distilled or deionized water.

(Please do not use tap water).

9. Strike the wells sharply onto absorbent paper or paper towels to remove all residual water droplets.

10. Dispense 200 μl of Enzyme Conjugate Reagent into each well. Gently mix for 10 seconds.

11. Incubate at 37°C for 30 minutes.

12. Remove the contents and wash the plate as described in step 7, 8, and 9.

13. Dispense 100 μl TMB Reagent into each well.


15. Incubate at room temperature in the dark for 20 minutes.

16. Stop the reaction by adding 100 μl of Stop Solution to each well.

17. Gently mix for 10 seconds. It is important to make sure that all the blue color changes to yellow color completely.

18. Read absorbance at 450nm with a microtiter well reader within 15 minutes.

CALCULATION OF RESULTS FOR SERUM AND PLASMA

1. Calculate the mean absorbance value (A450) for each set of reference standards, controls and patient samples.

2. Construct a standard curve by plotting the mean absorbance obtained from each reference standard against its concentration in μg/ml on graph paper, with absorbance values on the vertical or Y axis, and concentrations on the horizontal or X axis.

3. Use the mean absorbance values for each specimen to determine the corresponding concentration of B2MG in μg/ml from the standard curve.

ASSAY PROCEDURE FOR URINE TEST

1. Urine Samples need 10 fold Dilution with the Sample Diluent (i.e. 50 μl urine + 450 μl Sample Diluent).

2. Follow the same Assay Procedure for Serum/Plasma Test from step 2 to step 18.

CALCULATION OF RESULTS FOR URINE TEST

1. Calculate the mean absorbance value (A450) for each reference standards, controls and patient samples.

2. Construct a standard curve by plotting the mean absorbance obtained from each reference standard against its concentration in μg/ml on graph paper, with absorbance values on the vertical or Y axis, and concentrations on the horizontal or X axis.

3. Use the mean absorbance values for each specimen to determine the corresponding concentration of β2MG in μg/ml. Divide the calculated values by 10.1 (Since the β2 Microglobulin standards have been prediluted 101 fold, the results obtained from urine samples should be further divided by 10.1). For instance, if the calculated value for a urine sample from the standard curve is 2.40 μg/ml, then the real value will be 2.40 μg/ml ÷ 10.1 = 0.238 μg/ml/3.

LIMITATIONS OF THE PROCEDURE

1. Reliable and reproducible results will be obtained when the assay procedure is carried out with a complete understanding of the package insert instructions and with adherence to good laboratory practice.

2. The wash procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings.

3. Serum samples demonstrating gross lipemia, gross hemolysis, or turbidity should not be used with this test.

4. The results obtained from the use of this kit are for research use only and are not intended to be used as a part of any official diagnosis.

Example of a Standard Curve

<table>
<thead>
<tr>
<th>B2MG (μg/ml)</th>
<th>Absorbance (450 nm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>0.052</td>
</tr>
<tr>
<td>0.625</td>
<td>0.377</td>
</tr>
<tr>
<td>1.25</td>
<td>0.745</td>
</tr>
<tr>
<td>2.5</td>
<td>1.414</td>
</tr>
<tr>
<td>5.0</td>
<td>2.085</td>
</tr>
<tr>
<td>10.0</td>
<td>2.942</td>
</tr>
</tbody>
</table>

This standard curve is for the purpose of illustration only, and should not be used to calculate unknowns. Each user should obtain his or her own data and standard curve.